SHORT COMMUNICATION



Sunbathing on the southern beach: evidence of Egyptian mastigurs (*Uromastyx aegyptia*) on the Island of Tunb-e Bozorg, Persian Gulf

Hamzeh Oraie^{1, 2*}

¹Department of Zoology, Faculty of Science, Shahrekord University, Shahrekord, Iran ²Institute of Biotechnology, Shahrekord University, Shahrekord, Iran

Abstract

The herpetofauna of numerous Iranian islands in the Persian Gulf is largely unknown, and baseline data are urgently needed to monitor future trends. This study provides for the first time evidence that *Uromastyx aegyptia leptieni* is present on Tunb-e Bozorg Island in the Persian Gulf, Iran. Mitochondrial 16S rRNA sequence data were used to infer the phylogenetic relationship of the two Egyptian mastigurs from Tunb-e Bozorg Island. The dorsal color pattern of the Iranian specimens perfectly matches with the original description of *Uromastyx aegyptia leptieni*. In addition, our phylogenetic analysis shows that the examined specimens are identical to a specimen of *Uromastyx aegyptia leptieni* from the United Arab Emirates. The special conditions on Tunb-e Bozorg Island have provided a suitable habitat for *Uromastyx aegyptia leptieni*, which has led to a relatively large and self-sustaining population.

Received: 13 August 2024

Keywords: Iranian Islands, Persian Gulf, Uromastycinae, mtDNA, Haplotype, taxonomy

Accepted: 29 September 2024

The spiny-tailed lizards of the genus *Uromastyx* Merrem, 1820 are distributed in the arid regions of North Africa and extend across the Arabian Peninsula towards Iran (Sindaco and Jeremčenko, 2008; Uetz et al., 2023; Wilms, 2005; Wilms et al., 2009). The genus comprises a total of 15 species, of which *Uromastyx aegyptia* (Forskål, 1775) has been recorded in the coastal region of Iran (Uetz et al., 2023; Anderson 1999; Tamar et al., 2018). *Uromastyx aegyptia* (Forskål, 1775) is the largest species within the genus, with a potential body length of over 700 mm and a weight of up to 2,500 g (Wilms et al. 2009b). Three subspecies have been identified within this species: *Uromastyx aegyptia aegyptia* (Forskål, 1775), *Uromastyx aegyptia microlepis* Arnold 1980, and *Uromastyx aegyptia leptieni* Wilms & Böhme 2000 (Uetz et al., 2023). The species is currently classified as "Vulnerable" in the IUCN Red List of Threatened Species (IUCN, 2022) and has been listed in Appendix II of CITES (Safaei-Mahroo et al., 2015).

The Uromastyx aegyptia demonstrates remarkable adaptability to the extreme conditions of desert ecosystems, utilizing macro- and microhabitat shifting to facilitate effective thermoregulation (Wilms, 2005). It prefers to inhabit burrows with a length of up to 1025 cm and a depth of approximately 180 cm (Bouskila, 1983). Various environmental factors, such as climatic conditions, soil characteristics, vegetation density, and altitude, significantly influence the habitat preferences of *U. aegyptia* (Wilms et al., 2009b; Aghanajafizadeh and Mobaraki, 2018). It is noteworthy that *U. aegyptia* can adjust its dietary patterns, transitioning from a primarily herbivorous diet (based on a diverse array of native plant species) to a mainly carnivorous diet (consisting of insects and other arthropods) (e.g., Bouskila, 1987; Cunningham, 2000; Castilla et al., 2011a; Castilla et al., 2011b).

*Corresponding Author: 🖂 <u>stenodactylus@gmail.com</u>



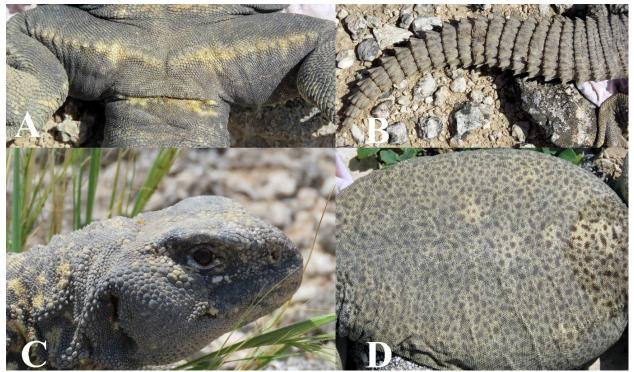


FIGURE 1. Photographs of *Uromastyx aegyptia leptieni*, caught on the island of Tunb-e Bozorg, Persian Gulf. Ventral view of the cloacal region, preanal and femoral pores present (A); whorls of spiny scales on the upper side of the tail (B); lateral view of the head (C); no enlarged scales on the upper side of the back (D).

Notwithstanding the absence of taxonomic studies, the Iranian populations are assigned to the subspecies *Uromastyx aegyptia microlepis* with a distribution range in the southern provinces of Hormozgan, Bushehr, and Fars (e.g., Anderson, 1999; Safaei-Mahroo et al., 2015). Herein, I present novel molecular data from two specimens collected from Tunb-e Bozorg Island, accompanied by natural history observations. It is noteworthy that Tunb-e Bozorg Island is one of the minor Iranian islands situated at the entrance of the Strait of Hormuz in the Persian Gulf, southern Iran.

During the field survey conducted on the Tunb-e Bozorg Island from May 6-9, 2024, aimed at examining the reptile's fauna, two juvenile specimens of *Uromastyx* were captured alive. The first step was to take high-resolution pictures of different body parts (Figure 1). Following standard protocols, blood samples were obtained from the caudal vein, and the animals were subsequently released back into their natural habitat (Figure 2B). The collected blood samples (HAC 1378, HAC 1379) were preserved at a temperature of -20 °C in a nucleic acid preservation buffer (NAP) in the Shahrekord University Herpetological Collection (HAC), Iran.

Total genomic DNA was extracted from the blood samples following standard extraction protocols (Green & Sambrook, 2012). Primers L16S, 5' - CGCCTGTTTATCAAAAACAT -3' and H16S, 5'- CCGGTCTGAACTCAGATCACG -3' were used to amplify a fragment of 16S rRNA gene (Kocher et al. 1989). The PCR protocol was as follows: Denaturation, 95 °C for 3 min; 36 cycles of 95 °C for 40 sec, primer annealing at 52 °C for 40 sec, and sequence elongation at 72 °C for 80 sec; 72 °C for 10 min; and subsequent storage at 4°C for 3 min. The amplification products were sequenced on an automated sequencer ABI 3730XL (GeneAzma Genetic Group, Isfahan, Iran) according to standard protocols.

A 494 base pair segment of the mitochondrial DNA sequences of the 16S rRNA gene was generated in this research and then deposited in the National Center for Biotechnology Information (NCBI) (PQ164784 (HAC 1378); PQ164783 (HAC 1379)). Sequences were edited in Bioedit version 7.0.0 (Hall 1999) and aligned with CLUSTAL W (Thompson et al. 1994). Additionally, available

sequences of 16S rRNA for *Uromastyx aegyptia* were downloaded from GenBank (Table S1). The sequences that were acquired were trimmed, finally, a dataset with a final sequence length of 486 nucleotides was generated. An unrooted phylogenetic network between 16S haplotypes of *Uromastyx aegyptia* was constructed utilizing the TCS network algorithm (Clement et al. 2002) implemented in Hapsolutely 0.2.2 (Vences et al. 2021) with the use of default parameters.

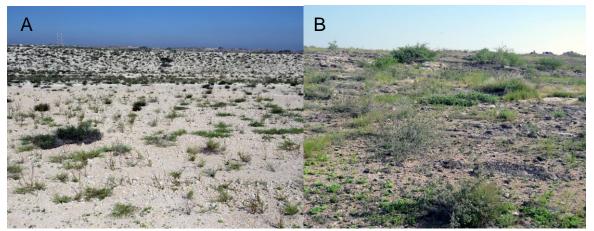


Figure 2. Different types of natural habitats of *Uromastyx aegyptia leptieni* on the island of Tunb-e Bozorg, Persian Gulf. () stony ground and gravelly plains with scattered bushes; (B) The flat silty gravel plains dominated by sparse vegetation.

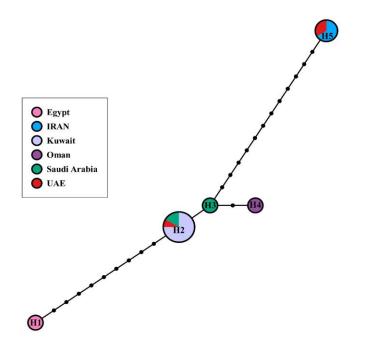


FIGURE 3. Phylogenetic network haplotypes based on for the 16S rRNA fragment (486 bp) in *Uromastyx* aegyptia. Circles correspond to haplotypes, with size proportional to the number of individuals per haplotype. Vertical lines correspond to mutational Colors steps. represent the geographic origins of haplotypes according to Table S1.

The analysis of the 16S rRNA sequences provides convincing evidence that the samples collected on Tunb-e Bozorg Island belong to *Uromastyx aegyptia leptieni* (Figure 3). In addition, the fact that the dorsal coloration pattern observed in the Iranian specimens is in perfect agreement with the original description of *Uromastyx aegyptia leptieni* (Figure 1). Unfortunately, the lack of the necessary equipment prevents the verification of the ventral count in the field (see Wilms & Böhme 2000). This study provides the first confirmation of the existence of *Uromastyx aegyptia leptieni* in Iran. Moreover, since the type locality of *Uromastyx aegyptia microlepis* (near Basrah, Iraq) is geographically adjacent to Iran, it is plausible that this subspecies also occurs on Iranian territory. To confirm or refute this hypothesis, comprehensive environmental and genetic studies are required.

On the island of Tunb-e Bozorg, my observations have shown that *Uromastyx aegyptia leptieni* inhabits two different natural habitats (Figure 2). One habitat, characterized by flat, silty gravel plains and sparse vegetation, hosted a significant population of *Uromastyx aegyptia leptieni* with over 40 active nests (Figure 2A). These lizards-built burrows with a single entrance and quickly retreated into them as soon as they noticed a potential threat, making capture in this environment seemingly impossible. Due to the extreme depth of these burrows, an enormous amount of energy is required to dig down to the ground. Nevertheless, these holes were expertly dug in a zigzag and spiral pattern. The second habitat consisted of stony ground and gravelly plains dotted with scattered bushes (Figure 2B), where the lizards resided in crevices under rocks, with no signs of burrow construction. Some researchers have claimed that the spiny-tailed lizard shows a preference for basking just after sunrise (Cunningham, 2000). However, my personal observations did not agree with this claim, as the lizards were predominantly observed either basking or foraging in the grass near their burrow entrances at around 8am. According to my observations, its sympatric and/or syntopic lizard and snake species include *Echis carinatus*, *Hemidactylus sp.*, *Pristurus sp.* and *Mesalina brevirostris*.

Globally, the Egyptian spiny lizard's habitat is shrinking due to factors such as overgrazing by livestock, human settlement, large-scale agricultural expansion, land reclamation, waste disposal and offroad vehicle traffic (IUCN, 2022). The expansion of military infrastructure on Tunb-e Bozorg Island has likely resulted in the loss of a significant proportion of suitable habitat for *Uromastyx aegyptia leptieni*. Despite these challenges, the Egyptian spiny lizard still thrives in some areas of Tunb-e Bozorg Island. Human-induced habitat degradation has contributed significantly to the increase in species extinction rates and the decline of populations worldwide (Burriel-Carranza et al., 2024). In certain countries, *Uromastyx aegyptia* is captured and traded for its skin to make leather, while its meat is used as a source of protein (e.g. Monchot et al., 2014; Aloufi et al., 2019). So far, no damage to the spiny-tailed lizard has been reported by people living on Tunb-e Bozorg Island.

In summary, the unique circumstances on Tunb-e Bozorg Island have created a favorable environment for *Uromastyx aegyptia leptieni* in some regions, resulting in a relatively large and selfsustaining population. There is evidence that military training areas not only fulfill their primary purpose of defense and training, but can also play an important role in environmental conservation by providing refuges for wildlife and helping to maintain healthy ecosystems (e.g. Pascaline and Sébastien, 2024). It appears that *Uromastyx aegyptia leptieni* is thriving on Tunb-e Bozorg Island without the need for a specific conservation program and that there is no immediate threat. However, it is important to point out that the two mitochondrial sequences used in this study are not sufficient to accurately assess the genetic diversity of this population. Therefore, I look forward to the possibility of using additional genetic markers in future studies to investigate the genetic diversity of *Uromastyx aegyptia* in the southern regions of Iran.

Acknowledgments

I express my deep gratitude to the Iranian Navy stationed on the Island of Tunb-e Bozorg for giving me the opportunity to explore the southernmost regions of my beloved country. My special thanks also go to Hamid Reza Mehdi-Abadi, who was doing his compulsory military service, for his unreserved help in my research expeditions in the particularly extremely hot and humid weather of Tunb-e Bozorg Island. I am also grateful for the partial financial support provided by the grant 02GRD1M31721 from Shahrekord University.

SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.

TABLE S1. Information of all sequences included in this study along with their localities and NCBI accession numbers.

DATA AVAILABILITY STATEMENT

The data for this study, including accession numbers for genetic sequences deposited on NCBI GenBank, are recorded in the Supporting Information, Table S1.

LITERATURE CITED

Aghanajafizadeh S, Mobaraki A. 2018. Habitat Selection by Spiny-tailed lizard (*Uromastyx aegyptia*) in Hengam Island, Iran. *The Saudi Journal of Life Sciences* 3: 414–419.

Aloufi AA, Amr ZS, Abu Baker MA, Hamidan N. 2019. Diversity and conservation of terrestrial, freshwater and marine reptiles and amphibians in Saudi Arabia. *Amphibian and Reptile Conservation* 13: 181–202.

Anderson SC. 1999. The Lizards of Iran. Society for the Study of Amphibians and Reptiles, Ithaca, New York.

Bouskila A. 1983. The burrows of the dabb-lizard, *Uromastyx aegyptius*. *Israel Journal of Zoology* 32: 151–152.

Bouskila A. 1987. Feeding in the herbivorous lizard *Uromastyx aegyptius* near Hazeva. *Israel Journal of Zoology* 33:122.

Burriel-Carranza B, Mochales-Riaño G, Talavera A, Els J, Estarellas M, Al Saadi S, Urriago Suarez JD, Olsson PO, Matschiner M, Carranza S. 2024. Clinging on the brink: Whole genomes reveal humaninduced population declines and severe inbreeding in the Critically Endangered Emirati Leaf-toed Gecko (*Asaccus caudivolvulus*). *Molecular Ecology* 33 :e17451.

Castilla AM, Richer R, Herrel A, Conkey AAT, Tribuna J, Chan R, Martínez de Aragón J, Böer B, Mohtar R. 2011a. Plant diversity in the diet of the lizard *Uromastyx aegyptia microlepis* in Qatar: The effect of zone, sampling date and faeces size. Proceedings of the Qatar Foundation Annual Research Forum 1, p. EVP7. 19

Castilla A, Richer R, Herrel A, Conkey A, Tribuna J, Al-Thani M. 2011b. First evidence of scavenging behaviour in the herbivorous lizard *Uromastyx aegyptia microlepis*. *Journal of Arid Environments* 75: 671–673.

Cunningham P. 2000. Daily activity pattern and diet of a population of the Spinytailed lizard, *Uromastyx aegyptius microlepis*, during summer in the United Arab Emirates. *Zoology in the Middle East* 21: 37–46.

Clement M, Snell Q, Walke P, Posada D, Crandall, K. 2002. TCS: estimating gene genealogies. Proc 16th Int Parallel Distrib Process Symp 2:184.

IUCN, 2022. The IUCN Red List of Threatened Species (Online). Available: https://www.iucnredlist.org. Accessed November 22th 2022.

Kocher TD, Thomas WK, Meyer A, Edwards SV, Pääbo S, Villablanca FX, Wilson AC. 1989. Dynamics of mitochondrial DNA evolution in animals: amplification and sequencing with conserved primers. *Proceedings of the National Academy of Sciences* 86: 6196–6200.

Hall TA. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic acids symposium series* 41: 95–98.

Monchot H, Bailon S, Schiettecatte J. 2014. Archaeozoological evidence for traditional consumption of spiny-tailed lizard (*Uromastyx aegyptia*) in Saudi Arabia. *Journal of Archaeological Science* 45: 96–102.

Pascaline C, Sébastien G. 2024. Contribution of military training areas for the conservation of calcareous grasslands, *Journal for Nature Conservation* 78:126579.

Safaei-Mahroo B, Ghaffari H, Fahimi H, Broomand S, Yazdanian M, Najafi Majd E, et al., 2015. The Herpetofauna of Iran: Checklist of Taxonomy, Distribution and Conservation Status. *Asian Herpetological Research* 6: 257–290.

Sindaco R, Jeremčenko VK. 2008. The reptiles of the Western Palearctic. 1. Annotated checklist and distributional atlas of the turtles, crocodiles, amphisbaenians and lizards of Europe, North Africa, Middle East and Central Asia. Edizioni Belvedere Latina, Monografie della Societas Herpetologica Italica.

Tamar K, Metallinou M, Wilms T, Schmitz A, Crochet P, Geniez P, Carranza S. 2018. Evolutionary history of spiny-tailed lizards (Agamidae: *Uromastyx*) from the Saharo-Arabian region. *Zoologica Scripta* 47:159–173.

Thompson JD, Higgins DG, Gibson TJ. 1994. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic acids research* 22: 4673–4680.

Uetz P, Freed P, Hzrošek J. 2023. The reptile database. Retrieved from http://www.reptile-database.org.

Vences M, Miralles A, Brouillet S, Ducasse J, Fedosov A, Kharchev V, Kumari S, Patmanidis S, Puillandre N, Scherz MD, Kostadinov I, Renner SS. 2021. iTaxoTools 0.1: Kickstarting a specimen-based software toolkit for taxonomists. *Megataxa* 6: 77–92.

https://www.mapress.com/mt/article/view/megataxa.6.2.1

Wilms T. 2005. Uromastyx – Natural History, Captive Care, Breeding. Offenbach, Germany: Herpeton.

Wilms T, Böhme W. 2000. A new *Uromastyx* species from south-eastern Arabia, with comments on the taxonomy of *Uromastyx aegyptia* (Forskål, 1775) (Squamata: Sauria. Agamidae). *Herpetozoa* 13: 133–148.

Wilms T, Böhme W, Wagner P, Lutzmann N, Schmitz A. 2009a. On the phylogeny and taxonomy of the genus *Uromastyx* Merrem, 1820 (Reptilia: Squamata: Agamidae: Uromastycinae): Resurrection of the genus *Saara* Gray, 1845. *Bonner Zoologische Beiträge*, 56, 55–99.

Wilms T, Wagner P, Shobrak M, Böhme W. 2009b. Activity profiles, habitat selection and seasonality of body weight in a population of Arabian spiny-tailed lizards (*Uromastyx aegyptia microlepis* Blanford, 1875; Sauria: Agamidae) in Saudi Arabia. *Bonner Zoologische Beiträge* 56: 259–272.