RESEARCH ARTICLE

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Haematology of the Levant Green Frog, *Pelophylax bedriagae* (Amphibia: Ranidae) in southern Iran

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Abstract

In this study, 68 specimens of the Levant Green Frog, *Pelophylax bedriagae*, were collected from agricultural fields around Yasouj and Noor-Abad cities, from August 2018 to May 2019 in southern Iran to investigate the blood cell morphology. Blood specimens were taken from the heart, or abdominal and facial veins. Smears were dried in room temperature, fixed by methanol and stained using Giemsa method. Blood cells (erythrocytes, leucocytes, and thrombocytes) were counted and measured using light microscopy under an optical micrometer. Based on the findings of this study, erythrocytes from the Noor-Abad population has significantly greater values relative to Yasouj population in all of the six metric characters (P < 0.05). RBC's in male frogs are also greater than those in females in all six studied characters (P < 0.05). All studied metric characters in lymphocytes are also different between two localities and sexes (P < 0.05).

Key words: RBC, Leucocyte, Yasouj, Noor-Abad, Haematology.

INTRODUCTION

Amphibians comprise 8090 known species with a worldwide distribution except for Antarctica and some oceanic Islands (AmphibiaWeb, 2019). The Levant Green Frog *Pelophylax bedriagae* (Camerano, 1882) is one of the three known representatives of the family Ranidae in Iran (AmphibiaWeb, 2019). Phylogenetic relationships of water frogs in Iran have studied by Pesarakloo et al. (2017), demonstrating two distinct mtDNA clades. The first clade, *Pelophylax bedriagae*, is primarily found from northwestern to southwestern parts of Iran. The second clade, which is different from other water frogs in genetic composition, distributes in the northeastern part of Iran.

Blood analysis is a useful method to determine the health of animals as well as managing and treating diseases (Christopher *et al.*, 1999). Multicellular animals need a circulatory system to carry nutrients and oxygen to and/or excretions from cells. Regulating blood composition and volume at a constant level is necessary for the continuity of animal life (Arikan & Çiçek, 2014). There is an increasing tendency toward the hematology of anurans during the recent decade (Davis *et al.*, 2008; Shutler & Marcogliese, 2011; Baraquet *et al.*, 2013; Arikan & Çiçek, 2014). Amphibian blood cells comprise erythrocytes, leukocytes, and thrombocytes (Arikan & Çiçek, 2014). Frog's erythrocytes are oval, nucleated, and biconcave, while the leucocytes look like those found in human blood (Storer *et al.*, 1957). Amphibians show well-developed circulatory and immune systems (Manning & Horton, 1982). Erythrocyte durability and longevity in amphibians are longer than those in birds and mammals. *Bufo marinus* RBC's, for instance, remain in blood circulation for

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around 700-1400 days (Altland & Brace, 1962). Leucocyte quality and quantity are regarded as an important hematologic parameter and a determinant of immune response level against diseases (Tierney et al., 2004), as when the number of eosinophils increases in blood parasite cases (Kiesecker, 2002). Leukocytes are divided into two groups based on the presence of granules in the cytoplasm and the shape of the nucleus: granulocytes (polymorphonuclear leukocytes) and agranulocytes (mononuclear leukocytes). Both of the two groups are spherical in blood plasma. Granulocytes comprise neutrophils, eosinophils, and basophils. The neutrophils are spherical cells with multiple nuclei connecting by thin threads of chromatin (Javanbakht et al., 2013). The eosinophils are the same size as neutrophils, having a doublet nucleus with many large granules in cytoplasm stained well by eosin (Javanbakht *et al.*, 2013). The basophils contain a unilobed nucleus, cytoplasmic granules, but differ in size from species to species (Wright, 2001). The basophils are predominant leucocytes in some amphibian species (Thrall *et al.*, 2012). Agranulocytes lack specific granules, but azurophiles granules and round denticulate nucleus, comprising monocyte and lymphocytes. As predominant leukocytes, small and large lymphocytes present in the blood of many amphibian species (Thrall *et al.*, 2012). Small lymphocytes are almost filled with the nucleus. Confusing generally with each other, monocytes are similar morphologically to large lymphocytes. The former is recognized from the latter by having a bean-like nucleus (Arikan & Çiçek, 2014). All vertebrate classes, except for mammals, are characterized by nucleated ervthrocytes. Unlike other vertebrates, the presence of a nucleus in mammalian erythrocytes is a pathologic symptom (Arserim & Mermer, 2008). Like all other blood cells in amphibians, thrombocytes are nucleated fusiform cells with the same function as mammalian platelets. Immature thrombocytes are spherical having a round nucleus and a more basophilic cytoplasm. These blood cells are generally found in clusters, making them more easily to be distinguished from small lymphocytes (Arıkan & Cicek, 2014). Given the fact that the hematology of the Levant Green Frog (Pelophyax bedriagae) has not yet been investigated in the southern parts of Iran, we try in the current survey to study and report some primary morphological parameters of blood cells of this species.

MATERIAL AND METHODS

This study carried out in two different localities in southern parts of Iran: Noor-Abad Mamasani (51° 34'E, 30°13'N; 900 masl) and Yasouj city (51° 34'E, 30°40'N; 1810 masl) in Fars and Kohgilouyeh & Boyer-Ahmad Provinces, respectively (Fig. 1). We collected 68 Pelophylax bedriagae with uneven distribution between the two localities including 43 (22 males, 21 females) and 25 (nine males, 16 females) specimens from Yasouj and Noor-Abad, respectively. These frogs were measured from snout to vent (SVL) using a digital caliper with an accuracy of 0.01 mm. We used the presence of vocal sacs in males as a distinguishing character to determine the sex of each Pelophylax bedriagae specimen (Bamezar et al., 2019). We took the blood from facial and abdominal veins as well as the heart in some specimens using insulin syringes. All the specimens were released immediately after blood taking. Two blood smears were prepared for each of the frogs, allowed to dry in ambient temperature for 10 minutes, fixed with absolute Methyl alcohol, and stained with the Giemsa method for 20 minutes. To study the morphology of blood cells, we analyzed ten different areas for each smear under 100X magnification using an Olympus CX31 microscope. Using an optical micrometer (BBT Krauss), four cellular characters measured in this study comprising cell length (CL), cell width (CW), nucleus length (NL), and nucleus width (NW) (Arıkan & Çiçek, 2014; Baraquet et al., 2013). To calculate the size of cells and nuclei, we used formulas LW $\pi/4$ for erythrocytes and A = πr^2 for leukocytes, respectively, in which: r = radius, L = length, and W = width (Arıkan & Çiçek, 2010; Javanbakht *et al.*, 2013).

Statistical analyses carried out using Excel (2016) and SPSS 19. Normality tests were carried out to choose between the appropriate parametric (t-Test) or non-parametric (Mann-Whitney *U*) tests between two groups (both localities and sexes) at the significance level of 0.05.

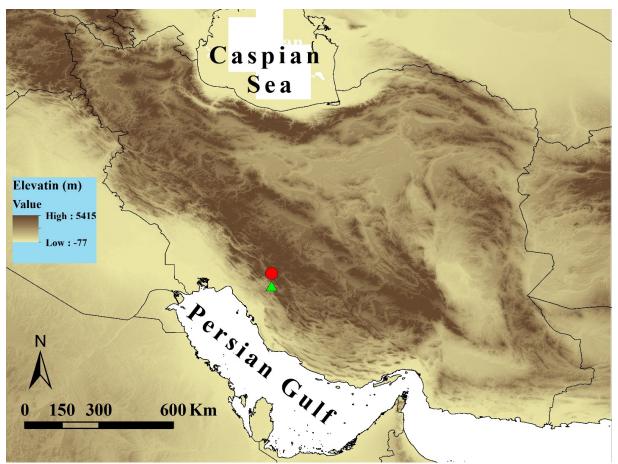


FIGURE 1. Position of collection sites of *Pelophylax bedriagae* in Noor-Abad Mamasani (green triangle) and Yasouj (red circle) in southern Iran.

RESULTS

Sex differences

The results of t-Test show that the mean value of SVL for females (71.51±11.53) is significantly larger than the males (62.02±7.24; P = 0.0001), while this value is not significantly different between two localities (i.e. Yasouj = 65.17±10.99 and Noor-Abad = 68.93±9.80) (P = 0.14).

Erythrocytes (RBCs, Fig. 2a)

A total number of 4032 RBCs were measured and analyzed in this study, of which 2533 cases were from Yasouj and 1499 from Noor-Abad regions, respectively. The values range from 17–28 μ m (average = 22.37) for RBC length, 9–17 μ m (average = 12.95) for RBC width, 6–12 μ m (average = 8.96) for nucleus length, and 4–7 μ m (average = 5.09) for nucleus width. All four characters showed non-normal distribution (Shapiro-Wilk test; *P* = 0.0001). The population from Noor-Ababd has significantly larger values than the Yasouj population for all the four characters (i.e., cell length, cell width, nucleus length, and nucleus width) (U-test; *P* = 0.0001; Table 1). Moreover, the Noor-Ababd population has larger values in both cell and nucleus sizes than the Yasouj population (U-test; *P* = 0.0001; Table 1). Male frogs have greater values for the four RBS's characters as well as cell and nucleus sizes than female frogs (U-test; *P* < 0.005; Table 2).

Neutrophils (Fig. 2b)

Only two out of six characters (nucleus length and cell width) were normally distributed (Shapiro-Wilk test; P > 0.05). The smallest and largest size of neutrophils is 86.35 and 379.16 μ m²,

respectively with an average of 181.70±58.62 μ m². There were no significant differences in all six characters between two sexes or localities (t-Test and U-test; P > 0.05; Tables 3-4).

Eosinophils (Fig. 2c)

Three out of six characters (i.e. nucleus length, cell length, and cell width) are normally distributed (Shapiro-Wilk test, P > 0.05). The average eosinophil size in both populations combined is 184.02±66.85 µm² ranging from 63.59 to 431.75 µm². Only one out of the six studied characters (i.e. cell size) are significantly different between Yasouj and Noor-Abad regions (*U*-test, *P* = 0.031; Table 5). Males and female frogs have the same values for all of the six studied characters (t-Test and U-test, *P* > 0.05; Table 6).

Basophils (Fig. 2d)

All of the three studied characters in basophils are normally distributed (Shapiro-Wilk test, P > 0.05). Basophil average size in both populations combined is 117.85±41.52 μ m² ranging from 37.68 to 240.21 μ m². There are no significant differences between the two groups (neither between two localities nor two genders) (t-Test, P > 0.05; Tables 7-8).

Lymphocytes (Fig. 2e)

All six relevant characters to lymphocytes show non-normal distribution (Shapiro-Wilk test, P = 0.0001). Lymphocyte average size in both populations combined is $126.17\pm56.84 \ \mu\text{m}^2$ ranging from 38.47 to 373.66 $\ \mu\text{m}^2$. The lymphocyte nucleus average size is 75.19 ± 30.54 ranging from 19.63 to 254.34 $\ \mu\text{m}^2$ (Table 9). All of the six studied characters are significantly different between to localities with larger values for Noor-Abad than Yasouj (U-test, P < 0.05; Table 9) as well as between two sexes with larger values for males than females (U-test, P < 0.05; Table 10).

Monocytes (Fig. 2f)

All six characters have a non-normal distribution (Shapiro-Wilk test; P < 0.05). Monocyte average size in both populations combined is $162.12\pm58.14 \ \mu\text{m}^2$ ranging from 78.50 to 373.66 $\ \mu\text{m}^2$. The monocyte nucleus average size is 81.35 ± 32.65 ranging from 28.26 to 200.96 $\ \mu\text{m}^2$ (Table 11). Only one out of the six studied characters (i.e. nucleus length) is significantly different between two localities with larger values for Noor-Abad than Yasouj (U-test, P = 0.033; Table 11) as well as between two sexes with larger values for males than females (U-test, P < 0.015; Table 12).

Thrombocytes (Fig. 2f)

All five characters studied in thrombocytes have a non-normal distribution (Shapiro-Wilk test; P < 0.05). Thrombocyte average size in both populations combined is $81.80\pm20.45 \ \mu\text{m}^2$ ranging from 47.10 to 138.1 $\ \mu\text{m}^2$ (Table 13). Three out of the five studied characters (i.e. nucleus width, cell width, and cell size) are significantly different between two localities with larger values for Noor-Abad than Yasouj (U-test, P < 0.05; Table 13). Thrombocyte cell width and size in male *Pelophylax bedriagae* have significantly larger values than females (U-test, P < 0.05; Table 14).

Characters	Locality	No.	Mean±SD	Min.	Max.	Range	Sig.	Test used
RBC N. Length	Yasouj	2533	8.69±0.96	5.00	13.00	8.00	0.0001	U-Test
	Noor-Abad	1499	9.40±1.01	5.00	13.00	8.00	-	
	Total	4032	8.90±1.14	5.00	13.00	8.00	-	
RBC N. Width	Yasouj	2533	4.96±0.74	4.00	8.00	4.00	0.0001	U-Test
	Noor-Abad	1499	5.33±0.70	4.00	8.00	4.00	-	
	Total	4032	5.10±0.75	4.00	8.00	4.00	_	
RBC Length	Yasouj	2533	21.70±1.91	15.00	29.00	14.00	0.0001	U-Test
	Noor-Abad	1499	23.50±2.14	17.00	33.00	16.00	-	
	Total	4032	22.38±2.20	15.00	33.00	18.00	_	
RBC Width	Yasouj	2533	12.55±1.31	8.00	19.00	11.00	0.0001	U-Test
	Noor-Abad	1499	13.63±1.54	9.00	19.00	10.00	-	
	Total	4032	12.95±1.49	8.00	19.00	11.00	_	
N. S.	Yasouj	2533	33.96±7.16	16.49	26.80	46.32	0.0001	U-Test
	Noor-Abad	1499	39.64±7.81	18.84	70.65	51.81	_	
	Total	4032	36.07±7.90	16.49	70.65	54.17	_	
RBC S.	Yasouj	2533	213.68±32.90	94.20	346.97	252.77	0.0001	U-Test
	Noor-Abad	1499	252.05±41.93	103.62	381.51	277.89	_	
	Total	4032	227.94±40.95	94.20	381.51	381.51	_	

TABLE 1. Descriptive statistics and significant analysis in erythrocytes of *Pelophylax bedriagae* between Yasouj and Noor-Abad regions in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

TABLE 2. Descriptive statistics and significant analysis in erythrocytes of male and female *Pelophylax bedriagae* in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

Characters	Sex	No.	Mean±SD	Min.	Max.	Range	Sig.	Test used
RBC N. Length	Male	2213	9.05±1.09	5.00	13.00	8.00	0.006	U-Test
	Female	1819	8.84±0.97	5.00	12.00	7.00	-	
	Total	4032	8.96±1.04	5.00	13.00	8.00	-	
RBC N. Width	Male	2213	5.20±0.80	4.00	8.00	0.30	0.0001	U-Test
	Female	1819	4.97±0.65	4.00	8.00	0.30	-	
	Total	4032	5.10±0.75	4.00	8.00	0.30	_	
RBC Length	Male	2213	22.80±2.35	16.00	33.00	1.10	0.0001	U-Test
	Female	1819	21.87±1.88	15.00	29.00	1.10	-	
	Total	4032	22.38±2.20	15.00	33.00	1.10	-	
RBC Width	Male	2213	13.10±1.52	9.00	19.00	0.80	0.0001	U-Test
	Female	1819	12.78±1.44	8.00	19.00	0.80	_	
	Total	4032	12.95±1.49	8.00	19.00	11.00	-	
Nucleus Size	Male	2213	37.24±8.58	16.49	69.08	52.60	0.0001	U-Test
	Female	1819	34.66±6.71	16.49	70.65	54.17	-	
	Total	4032	36.07±7.90	16.49	70.65	54.17	_	
RBC Size	Male	2213	235.03±43.61	94.20	372.88	278.68	0.0001	U-Test
	Female	1819	219.32±35.63	103.62	381.51	227.89	_	
	Total	4032	227.94±40.95	94.20	381.51	287.31	_	

Characters	Locality	No.	Mean±SD	Min	Max	Range	Sig.	Test used
Neutrophil N.	Yasouj	43	6.55±1.27	4.00	10.30	6.30	0.072	T- test
Length	Noor-Abad	19	7.13±1.32	4.50	9.70	5.20		
	Total	62	6.72±1.31	4.00	10.30	6.30		
Neutrophil N.	Yasouj	43	4.29±0.79	2.70	6.00	3.30	0.398	U- test
Width	Noor-Abad	19	4.47±0.87	2.30	6.30	4.00		
	Total	62	4.34±0.82	2.30	6.30	4.00		
Neutrophil	Yasouj	43	15.53±2.64	11.00	23.00	12.00	0.07	U- test
Length	Noor-Abad	19	16.89±2.49	14.00	22.00	8.00		
	Total	62	15.95±2.65	11.00	23.00	12.00		
Neutrophil	Yasouj	43	13.90±2.59	9.00	21.00	12.00	0.132	T- test
Width	Noor-Abad	19	14.94±2.14	11.00	19.00	8.00		
	Total	62	14.22±2.49	9.00	21.00	12.00		
Nucleus Size	Yasouj	43	24.44±10.26	9.62	68.38	58.76	0.11	U- test
	Noor-Abad	19	27.08±9.18	11.54	50.24	38.70		
	Total	62	25.25±9.94	9.62	68.38	58.76		
Neutrophil Size	Yasouj	43	174.59±59.11	86.55	379.94	293.39	0.051	U- test
	Noor-Abad	19	202.25±54.30	132.67	329.90	197.23		
	Total	62	183.07±58.66	86.55	379.94	293.39		

TABLE 3. Descriptive statistics and significant analysis in neutrophils of *Pelophylax bedriagae* between Yasouj and Noor-Abad regions in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

TABLE 4. Descriptive statistics and significant analysis in neutrophils of male and female *Pelophylax bedriagae* in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

Characters	Sex	No.	Mean±SD	Min	Max	Range	Sig.	Test used
Neutrophil N.	Male	29	6.88±1.39	4.30	10.30	6.00	0.526	T- test
Length	Female	33	6.58±1.23	4.00	9.00	5.00	_	
	Total	62	6.72±1.31	4.00	10.30	6.30	_	
Neutrophil N.	Male	29	4.38±0.94	2.30	6.30	4.00	0.782	U- test
Width	Female	33	4.30±0.70	2.70	6.00	3.30	_	
	Total	62	4.34±0.82	2.30	6.30	4.00	-	
Neutrophil	Male	29	15.72±2.69	11.00	22.00	11.00	0.545	U- test
Length	Female	33	16.15±2.64	11.00	23.00	12.00	_	
	Total	62	15.95±2.65	11.00	22.00	12.00	-	
Neutrophil	Male	29	13.86±2.40	10.00	18.00	8.00	0.286	T- test
Width	Female	33	14.54±2.57	9.00	21.00	12.00	-	
	Total	62	14.22±2.49	9.00	21.00	12.00	_	
Nucleus Size	Male	29	25.27±9.19	11.54	50.24	38.70	0.918	U- test
	Female	33	25.23±10.70	9.62	68.38	58.76	_	
	Total	62	25.25±9.94	9.62	68.38	58.76	_	
Neutrophil	Male	29	175.70±53.00	86.55	283.39	196.84	0.456	U- test
Size	Female	33	189.54±63.32	86.55	379.94	293.39	_	
	Total	62	183.07±58.66	86.55	379.94	293.39	_	

Characters	Locality	No.	Mean±SD	Min	Max	Range	Sig.	Test used
Eosinophil N.	Yasouj	38	9.21±1.78	5.50	13.00	7.50	0.748	T- test
Length	Noor-Abad	16	9.37±1.50	6.50	13.00	6.50		
	Total	54	9.26±1.70	5.50	13.00	7.50	_	
Eosinophil N.	Yasouj	38	5.41±1.14	3.50	9.00	5.50	0.946	U- test
Width	Noor-Abad	16	5.31±0.96	3.50	7.00	3.50	_	
	Total	54	5.37±1.08	3.50	9.00	5.50	_	
Eosinophil	Yasouj	38	15.47±3.28	9.00	25.00	16.00	0.107	T- test
Length	Noor-Abad	16	16.62±1.82	13.00	20.00	7.00	_	
	Total	54	15.81±2.95	9.00	25.00	16.00	_	
Eosinophil	Yasouj	38	14.02±2.75	9.00	22.00	13.00	0.09	T- test
Width	Noor-Abad	16	15.31±1.66	13.00	18.00	5.00	_	
	Total	54	14.40±2.53	9.00	22.00	13.00	_	
Nucleus Size	Yasouj	38	43.14±15.38	23.75	94.99	71.24	0.690	U-test
	Noor-Abad	16	42.89±10.07	23.75	67.17	43.42	_	
	Total	54	43.07±13.92	23.75	94.99	71.24	_	
Eosinophil	Yasouj	38	177.52.33±75.22	63.59	433.52	369.93	0.031	U- test
lize	Noor-Abad	16	201.92±39.06	132.67	268.67	136.00	_	
	Total	54	184.75±67.14	63.59	433.52	369.93	-	

TABLE 5. Descriptive statistics and significant analysis in eosinophils of *Pelophylax bedriagae* between Yasouj and Noor-Abad regions in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

TABLE 6. Descriptive statistics and significant analysis in eosinophils of male and female *Pelophylax bedriagae* in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

Characters	Sex	No.	Mean±SD	Min.	Max	Range	Sig.	Test used
Eosinophil	Male	20	8.75 ±1.62	6.50	12.00	5.50	0.091	T- test
N. Length	Female	34	9.55±1.68	5.50	13.00	7.50		
	Total	54	9.33±2.70	5.50	13.00	7.50		
Eosinophil	Male	20	5.42±1.06	3.50	7.00	3.50	0.536	U- test
N. Width	Female	34	5.32±1.11	3.50	9.00	5.50		
	Total	54	5.42±1.07	3.50	9.00	5.50		
Eosinophil	Male	20	15.25±2.75	9.00	19.00	10.00	0.286	T- test
Length	Female	34	16.14±3.06	12.00	25.00	13.00		
	Total	54	15.75±2.80	9.00	25.00	16.00		
Eosinophil	Male	20	14.00±2.42	9.00	18.00	9.00	0.370	T- test
Width	Female	34	14.64±2.60	10.00	22.00	12.00		
	Total	54	14.37±2.43	9.00	22.00	13.00		
Nucleus	Male	20	40.44±13.45	23.75	67.17	43.42	0.232	U- test
Size	Female	34	44.61±14.17	23.75	94.99	71.24		
	Total	54	43.07±13.92	23.75	94.99	71.24		
Eosinophil	Male	20	172.49±55.87	63.59	268.67	205.08	0.489	U- test
Size	Female	34	191.97±72.79	103.82	433.52	329.70		
	Total	54	184.75±67.14	63.59	433.52	369.93		

and	d Noor-Abad	regions in so	uthern	Iran. Length a	ind width in	µm and size	e in µm². Ab	breviation	s: N., nucleus;
No	., number; Ma	ax., maximum	; Min.,	minimum; SD,	standard de	eviation; Sig.	, significanc	e.	
-	Characters	Locality	No.	Mean±SD	Min	Max	Range	Sig.	Test
_									used
	Basophil	Yasouj	41	13.41±2.94	8.00	21.00	13.00	0.414	T- test
	Length	Noor-Abad	6	12.33±3.38	9.00	17.00	8.00		

8.00

6.00

9.00

6.00

38.47

63.59

38.47

21.00

17.00

15.00

17.00

240.41

188.60

240.41

13.00

11.00

6.00

11.00

201.94

125.01

201.94

0.945

0.714

T- test

T-test

TABLE 7. Descriptive statistics and significant analysis in basophils of *Pelophylax bedriagae* between Yasouj

TABLE 8. Descriptive statistics and significant analysis in basophils of male and female *Pelophylax bedriagae* in southern Iran. Length and width in µm and size in µm². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

Characters	Sex	No.	Mean±SD	Min.	Max.	Range	Sig.	Test used
Basophil Length	Male	14	12.28±2.75	8.00	17.00	9.00	0.141	T- test
	Female	33	13.69±3.02	8.00	21.00	13.00	_	
	Total	47	13.27±2.99	8.00	21.00	13.00	_	
Basophil Width	Male	14	10.57±2.24	8.00	15.00	7.00	0.291	T- test
	Female	33	11.33±2.23	6.00	17.00	11.00	-	
	Total	47	11.10±2.23	6.00	17.00	11.00	_	
Basophil Size	Male	14	106.31±42.34	50.24	188.60	138.36	0.149	T- test
	Female	33	126.38±42.13	38.47	240.41	201.94	_	
	Total	47	120.40±42.75	38.47	240.41	201.94	_	

TABLE 9. Descriptive statistics and significant analysis in lymphocytes of *Pelophylax bedriagae* between Yasouj and Noor-Abad regions in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

Characters	Locality	No.	Mean±SD	Min	Max	Range	Sig.	Test used
Lymphocyte	Yasouj	406	10.33±2.38	6.00	19.00	13.00	0.007	U- test
N. Length	Noor-Abad	298	10.75±2.19	5.00	19.00	14.00		
	Total	704	10.51±2.31	5.00	19.00	14.00		
Lymphocyte	Yasouj	406	8.51±1.92	4.00	17.00	13.00	0.014	U- test
N. Width	Noor-Abad	298	8.89±1.90	5.00	17.00	12.00	_	
	Total	704	8.67±1.92	4.00	17.00	13.00		
Lymphocyte	Yasouj	406	13.01±3.14	4.00	28.00	24.00	0.0001	U-Test
Length	Noor-Abad	298	14.36±3.82	7.00	27.00	20.00	_	
	Total	704	13.60±3.51	4.00	28.00	24.00		
Lymphocyte	Yasouj	406	11.02±2.38	6.00	21.00	15.00	0.0001	U- test
Width	Noor-Abad	298	11.78±2.62	4.00	19.00	15.00		
	Total	704	11.35±2.51	4.00	21.00	17.00		
Nucleus Size	Yasouj	406	72.74±30.78	8.55	226.87	218.32	0.007	U- test
	Noor-Abad	298	78.45±30.07	19.63	254.34	234.71		
	Total	704	75.16±30.59	8.55	254.34	254.79	_	
Lymphocyte	Yasouj	406	118.94±51.92	38.47	397.41	358.94	0.0001	U- test
Size	Noor-Abad	298	141.23±64.81	38.47	362.87	324.40	_	
	Total	704	128.38±58.73	38.47	397.41	358.94	_	

Basophil

Basophil

Size

Width

Total

Yasouj

Total

Yasouj

Total

Noor-Abad

Noor-Abad

47

41

6

47

41

6

47

13.27±2.99

 11.09 ± 2.25

11.16±2.31

11.10±2.23

121.5±41.95

113.00±51.55

120.4±42.75

Characters	Sex	No.	Mean±SD	Min	Max	Range	Sig.	Test used
Lymphocyte	Male	385	10.78±2.24	5.00	19.00	14.00	0.0001	U-Test
N. Length	Female	319	10.19±2.37	6.00	19.00	13.00		
	Total	704	10.51±2.32	5.00	19.00	14.00		
Lymphocyte	Male	385	8.80±1.92	4.00	17.00	13.00	0.041	U-Test
N. Width	Female	319	8.52±1.92	5.00	17.00	12.00		
	Total	704	8.67±1.92	4.00	17.00	13.00		
Lymphocyte	Male	385	14.26±3.70	7.00	28.00	21.00	0.0001	U-Test
Length	Female	319	12.84±3.05	7.00	23.00	16.00		
	Total	704	13.62±3.49	7.00	28.00	21.00		
Lymphocyte	Male	385	11.54±2.47	4.00	19.00	15.00	0.012	U-Test
Width	Female	319	11.13±2.56	6.00	21.00	15.00		
	Total	704	11.35±2.52	4.00	21.00	17.00		
Nucleus	Male	385	77.98±29.58	19.63	254.34	234.71	0.001	U-Test
Size	Female	319	71.82±31.38	23.75	226.87	203.12		
	Total	704	75.19±30.54	19.63	254.34	234.71		
Lymphocyte	Male	385	136.72±60.50	38.47	397.41	358.94	0.0001	U-Test
Size	Female	319	118.31±54.94	38.47	362.87	324.40		
	Total	704	128.38±58.73	38.47	397.41	358.94		

TABLE 10. Descriptive statistics and significant analysis in lymphocytes of male and female *Pelophylax bedriagae* in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

TABLE 11. Descriptive statistics and significant analysis in monocytes of *Pelophylax bedriagae* between Yasouj and Noor-Abad regions in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

Characters	Locality	No.	Mean±SD	Min	Max	Range	Sig.	Test used
Monocyte	Yasouj	36	11.86±2.72	8.00	20.00	12.00	0.033	U- test
N. Length	Noor-Abad	18	13.50±3.24	9.00	24.00	15.00		
	Total	54	12.41±2.97	8.00	24.00	16.00		
Monocyte	Yasouj	36	7.86±2.26	3.00	12.00	9.00	0.282	U- test
N. Width	Noor-Abad	18	7.00±2.52	3.00	11.00	8.00		
	Total	54	7.57±2.36	3.00	12.00	9.00		
Monocyte	Yasouj	36	15.81±3.45	11.00	28.00	17.00	0.321	U- test
Length	Noor-Abad	18	14.44±2.18	10.00	18.00	8.00		
	Total	54	15.35±3.13	10.00	28.00	18.00		
Monocyte	Yasouj	36	13.31±2.27	10.00	19.00	9.00	0.486	U- test
Width	Noor-Abad	18	12.83±2.18	10.00	18.00	8.00		
	Total	54	13.15±2.23	10.00	19.00	9.00		
Nucleus	Yasouj	36	79.62±34.15	28.26	200.96	172.70	0.586	U- test
Size	Noor-Abad	18	84.82±30.05	38.47	165.05	126.58		
	Total	54	81.35±32.65	28.26	200.96	172.70		
Monocyte	Yasouj	36	171.53±66.07	94.99	397.41	302.42	0.314	U- test
Size	Noor-Abad	18	149.16±44.70	78.50	254.34	175.84		
	Total	54	164.07±60.31	78.50	397.41	318.91		

Characters	Sex	No.	Mean±SD	Min	Max	Range	Sig.	Test used
Monocyte	Male	21	13.57±3.54	8.00	24.00	16.00	0.015	U- test
N. Length	Female	33	11.67±2.31	8.00	18.00	10.00		
	Total	54	12.41±2.97	8.00	24.00	16.00		
Monocyte	Male	21	7.24±2.19	4.00	12.00	8.00	0.324	U- test
N. Width	Female	33	7.79±2.47	3.00	12.00	9.00		
	Total	54	7.57±2.36	3.00	12.00	9.00		
Monocyte	Male	21	16.19±3.60	11.00	28.00	17.00	0.073	U- test
Length	Female	33	14.82±2.71	10.00	22.00	12.00		
	Total	54	15.35±3.13	10.00	28.00	18.00		
Monocyte	Male	21	13.78±2.27	10.00	18.00	8.00	0.327	U- test
Width	Female	33	12.94±2.21	10.00	19.00	9.00		
	Total	54	13.15±2.23	10.00	19.00	9.00		
Nucleus	Male	21	8.75±39.38	28.26	200.96	172.70	0.358	U-test
Size	Female	33	76.64±27.15	28.26	165.05	136.79		
	Total	54	81.35±32.65	28.26	200.96	172.70		
Monocyte	Male	21	178.02±68.23	94.99	397.41	302.42	0.141	U- test
Size	Female	33	155.20±53.90	78.50	298.50	220.00		
	Total	54	164.01±60.31	78.50	397.41	318.91		

TABLE 12. Descriptive statistics and significant analysis in monocytes of male and female *Pelophylax bedriagae* in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

TABLE 13. Descriptive statistics and significant analysis in thrombocytes of *Pelophylax bedriagae* between Yasouj and Noor-Abad regions in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

Characters	Locality	No.	Mean±SD	Min	Max	Range	Sig.	Test used
Thrombocyte N.Length	Yasouj	173	11.50±1.83	7.00	16.00	9.00	0.065	U- test
	Noor-	42	12.09±1.60	10.00	15.00	5.00		
	Abad						_	
	Total	215	11.62±1.80	7.00	16.00	9.00	-	
Thrombocyte N.Width	Yasouj	173	5.25±0.92	4.00	7.00	3.00	0.0001	U- test
	Noor- Abad	42	6.09±0.84	5.00	8.00	3.00	_	
	Total	215	5.41±0.96	4.00	8.00	4.00		
Thrombocyte Length	Yasouj	173	15.77±2.89	10.00	22.00	12.00	0.615	U- test
	Noor-	42	15.47±2.17	12.00	22.00	10.00		
	Abad						_	
	Total	215	15.71±2.76	10.00	22.00	12.00		
Thrombocyte Width	Yasouj	173	6.41±1.00	5.00	8.00	3.00	0.0001	U- test
	Noor- Abad	42	7.42±0.63	6.00	8.00	2.00	-	
	Total	215	6.61±1.02	5.00	8.00	3.00	_	
Thrombocyte Size	Yasouj	173	79.69±20.83	47.10	138.10	91.06	0.001	U- test
	Noor- Abad	42	90.46±16.37	65.94	138.10	72.22		
	Total	215	81.80±20.45	47.10	138.10	91.06		

Characters	Sex	No.	Mean±SD	Min	Max	Range	Sig.	Test used
Thrombocyte N. Length	Male	103	11.77±1.89	8.00	15.00	7.00	0.261	U- test
	Female	112	11.48±1.71	7.00	16.00	9.00		
	Total	215	11.62±1.80	7.00	16.00	9.00	-	
Thrombocyte N. Width	Male	103	5.52±0.95	4.00	8.00	4.00	0.184	U- test
	Female	112	5.32±0.96	4.00	7.00	3.00	_	
	Total	215	5.41±0.96	4.00	8.00	4.00		
Thrombocyte Length	Male	103	15.99±2.98	10.00	22.00	12.00	0.150	U- test
	Female	112	15.46±2.53	10.00	22.00	12.00		
	Total	215	15.71±2.76	10.00	22.00	12.00		
Thrombocyte Width	Male	103	6.82±1.00	5.00	8.00	3.00	0.004	U- test
	Female	112	6.41±1.01	5.00	8.00	3.00	-	
	Total	215	6.61±1.02	5.00	8.00	3.00		
Thrombocyte Size	Male	103	85.97±21.60	47.10	138.16	91.06	0.007	U- test
	Female	112	77.96±18.62	47.10	138.16	91.06		
	Total	215	81.80±20.45	47.10	138.16	91.06	-	

TABLE 14. Descriptive statistics and significant analysis in thrombocytes of male and female *Pelophylax bedriagae* in southern Iran. Length and width in μ m and size in μ m². Abbreviations: N., nucleus; No., number; Max., maximum; Min., minimum; SD, standard deviation; Sig., significance.

DISCUSSION

Stained with Giemsa, erythrocytes show a blue homogenous cytoplasm. Adult erythrocytes have a basophilic nucleus (Javanbakht *et al.*, 2013). Carrying O2 and CO_2 is one of the most important functions performing by RBCs. The ratio of erythrocyte surface area to size is regarded as a determining factor in tissues, with a higher rate of gas and nutrient exchange for smaller erythrocytes than larger ones. Erythrocyte size reflects the position of a species on an evolutionary scale. For instance, lower vertebrates and animal groups, which do not show a success evolutionary past (i.e. Cyclostomes, Elasmobranches, and Urodela), possess larger erythrocytes unlike smaller enucleated erythrocytes in the higher vertebrates such as mammals (Hartman & Lessler, 1964; Szarski & Czopek, 1966; Saint Girons, 1970). Noor-Abad population lies at a lower altitude than the Yasouj population (900 vs. 1810 masl). The results of this study show that the size of erythrocytes between two localities is significantly different, with larger average values in all six studied characters for the Noor-Abad than the Yasouj population. Some other studies have shown the blood cell size differences between different populations of the same species inhabiting different altitudes. For instance, the size of erythrocytes and their nuclei in Hypsiboas cordobae decreased significantly with increasing altitude (Baraquet et al., 2013). The smaller but higher number of erythrocytes improve the uptake of oxygen; finally adapt the organism to the environments having low oxygen pressures (Hutchison *et al.*, 1976). Due to higher ratio of surface area to volume and subsequently more efficiency in gas exchanges for smaller erythrocytes (Ruiz et al. 1983; Arıkan, 1989; Weber, 2007), possessing smaller erythrocytes might be regarded as an advantage for frogs inhabiting higher altitudes with lower oxygen pressure (like Yasouj population in this study) than lower altitudes with higher oxygen pressure (i.e. Noor-Abad population), explaining size differences between the two different localities. The same negative correlation between erythrocyte size and altitude has been reported in other amphibians (Ruiz et al. 1983; Arıkan, 1989; Weber, 2007; Baraquet *et al.*, 2013). Like other amphibians, frogs have oval erythrocytes encompassing an oval central nucleus. There is a negative correlation between the rate of metabolism and erythrocyte size, in that, more active species have smaller erythrocytes while the ones with less oxygen consumption have larger erythrocytes (Smith, 1925; Evans, 1939; Vernberg, 1955; Monnickendam & Balls, 1973).

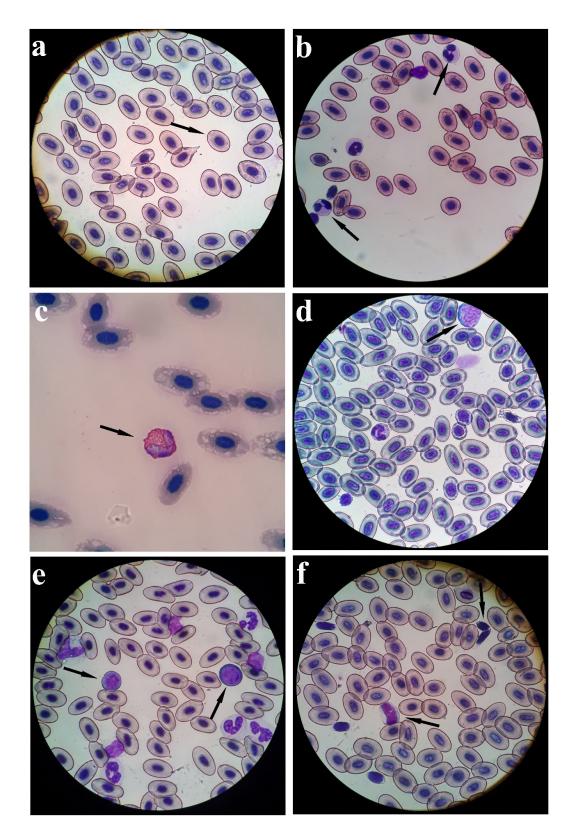


FIGURE 2. Blood cells in the peripheral circulatory system of *Pelophylax bedriagae*, including erythrocytes (a), neutrophils (b), eosinophil (c), basophil (d), lymphocytes (e), monocyte (f; lower arrow) and thrombocytes (f, upper arrow), under 100X magnification for (c) and 40X for the remaining visual fields.

Male-biased differences were also observed in all six characters studied in erythrocytes. The largest and most oval erythrocytes in anuran species have previously reported for aquatic *Pelophylax caralitanus*, while the smallest ones have reported in the terrestrial *Pelodytes caucasicus* (Atatür *et al.*, 1999; Arıkan *et al.*, 2001; Arikan *et al.*, 2003; Dönmez *et al.*, 2009; Erişmiş *et al.*, 2010; Arikan & Çiçek, 2014). The Farmed Bullfrog, *Lithobates catesbeianus*, have been shown to have larger erythrocyte morphometric parameters (i.e., surface area, the volume of RBCs and their nuclei) during active than hibernating period (Peng *et al.*, 2016). The erythrocyte size influenced by numerous factors including size of the animals (Vernberg, 1955), environmental conditions like temperature and atmospheric pressure (Ruiz *et al.*, 1983, 1989), and other conditions such as health status, reproduction, hibernation, feeding, and daily activities (Wojtaszek *et al.*, 1997; Allender & Fry, 2008; Thrall *et al.*, 2012).

Leukocytes are specialized blood cells that act primarily in specific and unspecific immunological responses (Iwama and Nakanishi, 1996). Leukocyte numbers differ significantly between active and hibernating seasons in the Farmed Bullfrog, Lithobates catesbeianus (Peng et al., 2016). Several factors determine the levels of leukocyte in the whole blood including environmental quality (LeaMaster et al., 1990), nutritional state (Barros et al., 2002), the presence of infectious agents (Martins et al., 2008), and parasitism (Martins et al., 2004; Arikan, 1989). Parasitic infections, for example, increase the number of monocytes and other leukocytes (Tavares-Dias et al., 2007). Analyses of the leukocytes in this study indicated that six characters of lymphocytes are significantly different between two localities as well as sexes. Only one out of six characters (nucleus length) in monocytes is different between the two localities and sexes, while the remaining characters of monocytes and all the characters in other leukocytes are the same between localities and sexes. This study showed that lymphocytes are the most abundant leukocvtes in the peripheral blood circulation of *Pelophylax bedriagae* which is consistent with the results of other studies conducted on anurans, say, Farmed Bullfrog, *Lithobates catesbeianus* (Peng et al., 2016), Dubio's Tree Frog, Polypedates teraiensis (Das & Mahapatra, 2014), American Bullfrog Rana catesbeiana (Cathers et al., 1997), and various species of frogs in Turkey (Arikan & Cicek, 2010), suggesting the primary role of immunological responses for lymphocytes among other leukocytes (Peng *et al.*, 2016). On the other hand, basophils have the smallest number of leukocytes in *Pelophylax bedriagae*, which is accordant with Dubio's Tree Frog *Polypedates teraiensis* (Das & Mahapatra, 2014) and Farmed Bullfrog *Lithobates catesbeianus* (Peng *et al.*, 2016). The basophil abundance in peripheral blood circulation depends on some factors such as species, season, geographic location, age of the animal, or probably blood parasites or viral infections (Vasaruchapong et al., 2014). Like other studies (Claver & Quaglia, 2009; Arıkan & Çiçek, 2010), a high concentration of granules in eosinophils and basophils, prevented us to observe and investigate nuclei of the two leukocytes.

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LITERATURE CITED

Allender, M.C., Fry, M.M., 2008. Amphibian hematology. Veterinary Clinics of North America: Exotic Animal Practice 11(3), 463–480.

Altland, P.D., Brace, K.C., 1962. Red cell life span in the turtle and toad. American Journal of Physiology-Legacy Content 203(6), 1188–1190.

AmphibiaWeb. 2019. https://amphibiaweb.org>, University of California, Berkeley, CA, USA, 6 Sep 2019.

Arıkan, H., Çiçek, K., 2010. Morphology of peripheral blood cells from various species of Turkish Herpetofauna. Acta Herpetologica 5(2), 179–198.

Arikan, H., Çiçek, K., 2014. Haematology of amphibians and reptiles: A review. North-Western Journal of Zoology 10(1), 190–209.

Arıkan, H., 1989. Anadolu'daki *Rana ridibunda* (Anura: Ranidae) populasyonlarının kan hücrelerinin sayısı bakımından incelenmesi. Turkish Journal of Zoology 13, 54–59.

Arikan, H., Atatür, M.K., Tosunoğlu, M., 2003. A study on the blood cells of the Caucasus Frog, *Pelodytes caucasicus*. Zoology in the Middle East 30(1), 43–47.

Arıkan, H., Çevik, İ.E., Kaya, U., Mermer, A., 2001. Anadolu Dağ Kurbağalarında Eritrosit Ölçümleri.

Arserim, S.K., Mermer, A., 2008. Hematology of the Uludağ Frog, *Rana macrocnemis* Boulenger, 1885 in Uludağ National Park (Bursa, Turkey). Su Ürünleri Dergisi 25(1), 39–46.

Atatür, M.K., Arikan, H., Çevik, I.E., 1999. Erythrocyte sizes of some anurans from Turkey. Turkish Journal of Zoology 23(2), 111–114.

Bamezar, F., Fathinha, B., Shafaeipour, A., 2019. Trophology of Levant Green Frog, *Pelophylax bedriagae* (Amphibia: Anura: Ranidae) in Choram Township, Kohgilouyeh & Boyer-Ahmad Province, Iran. North-western Journal of Zoology 15(2), 168–174.

Baraquet, M., Grenat, P.R., Salas, N.E., Martino, A.L., 2013. Intraspecific variation in erythrocyte sizes among populations of *Hypsiboas cordobae* (Anura, Hylidae). Acta Herpetologica 8(2), 93–97.

Barros, M.M., Pezzato, L.E., Kleemann, G.K., Hisano, H., Rosa, G.J.d.M., 2002. Níveis de vitamina C e ferro para tilápia do Nilo (*Oreochromis niloticus*). Revista Brasileira de Zootecnia 31(6), 2149–2156.

Cathers, T., Lewbart, G.A., Correa, M., Stevens, J.B., 1997. Serum chemistry and hematology values for anesthetized American bullfrogs (*Rana catesbeiana*). Journal of Zoo and Wildlife Medicine 171–174.

Christopher, M.M., Berry, K.H., Wallis, I., Nagy, K., Henen, B., Peterson, C., 1999. Reference intervals and physiologic alterations in hematologic and biochemical values of free-ranging desert tortoises in the Mojave Desert. Journal of wildlife diseases 35(2), 212–238.

Claver, J.A., Quaglia, A.I., 2009. Comparative morphology, development, and function of blood cells in nonmammalian vertebrates. Journal of exotic pet medicine 18(2), 87–97.

Das, M., Mahapatra, P.K., 2014. Hematology of wild-caught Dubois's tree frog *Polypedates teraiensis*, Dubois, 1986 (Anura:Rhacophoridae). The Scientific World Journal https://doi.org/10.1155/2014/491415.

Davis, A., Maney, D., Maerz, J., 2008. The use of leukocyte profiles to measure stress in vertebrates: a review for ecologists. Functional Ecology 22(5), 760–772.

Dönmez, F., Tosunoğlu, M., Gül, Ç., 2009. Hematological values in hermaphrodite, *Bufo bufo* (Linnaeus, 1758). North-Western Journal of Zoology 5(1), 97–103.

Erişmiş, U., Arikan, H., Alpagut-Keskin, N., Çevik, İ.E., 2010. A study on the blood cells of the firebellied toad, *Bombina bombina* L.(Anura: Bombinatoridae). Animal biology 60(1), 61–68.

Evans, G., 1939. Factors influencing the oxygen consumption of several species of plethodontid salamanders in aerial and aquatic media. Ecology 74–95.

Hartman, F., Lessler, M., 1964. Erythrocyte measurements in fishes amphibia, and reptiles. The Biological Bulletin 126(1), 83–88.

Hutchison, V.H., Haines, H.B., Engbretson, G., 1976. Aquatic life at high altitude: respiratory adaptations in the Lake Titicaca frog, *Telmatobius culeus*. Respiration physiology 27(1), 115–129.

Iwama, G., Nakanishi, T., 1996. *The immune system*. California: Academic Press. 380p.

Javanbakht, H., Vaissi, S., Parto, P., 2013. The morphological characterization of the blood cells in the three species of turtle and tortoise in Iran. Research in Zoology 3(1), 38–44.

Kiesecker, J.M., 2002. Synergism between trematode infection and pesticide exposure: a link to amphibian limb deformities in nature? Proceedings of the National Academy of Sciences 99(15), 9900–9904.

LeaMaster, B.R., Brock, J.A., Fujioka, R.S., Nakamura, R.M., 1990. Hematologic and blood chemistry values for *Sarotherodon melanotheron* and a red hybrid tilapia in freshwater and seawater. Comparative Biochemistry and Physiology Part A: Physiology 97(4), 525–529.

Manning M.J., Horton J.D., 1982. RES Structure and Function of the Amphibia. In: Cohen N., Sigel M.M. (eds) Phylogeny and Ontogeny. Springer, Boston, MA.

Martins, M., Mouriño, J., Amaral, G., Vieira, F., Dotta, G., Jatobá, A., Pedrotti, F., Jerônimo, G., Buglione-Neto, C., 2008. Alterações hematológicas em tilápia do Nilo infectada experimentalmente com Enterococcus sp. Brazilian Journal of Biology 68(3), 657–661.

Martins, M., Tavares-Dias, M., Fujimoto, R., Onaka, E., Nomura, D., 2004. Haematological alterations of *Leporinus macrocephalus* (Osteichtyes: Anostomidae) naturally infected by *Goezia leporini* (Nematoda: Anisakidae) in fish pond. Arquivo Brasileiro de Medicina Veterinária e Zootecnia 56(5), 640–646.

Monnickendam, M.A., Balls, M., 1973. The relationship between cell sizes, respiration rates and survival of amphibian tissues in long-term organ cultures. Comparative Biochemistry and Physiology Part A: Physiology 44(3), 871–880.

Peng, F., Zhang, R., Zhu, X., Wang, H., Zhang, S., 2016. Hematology and serum biochemistry of farmed bullfrog, *Lithobates catesbeianus* during the active and hibernating periods. Journal of Veterinary Medicine and Animal Health 8(11), 176–182.

Pesarakloo, A., Rastegar-Poyani, E., Rastegar-Poyani, N., Kami, H., Najibzadeh, M., Khosravani, A., Oraie, H., 2017. The first taxonomic revaluation of the Iranian water frogs of the genus *Pelophylax*

(Anura: Ranidae) using sequences of the mitochondrial genome. Mitochondrial DNA 28(3), 392–398.

Ruiz, G., Rosenmann, M., Veloso, A., 1983. Respiratory and hematological adaptations to high altitude in *Telmatobius* frogs from the Chilean Andes. Comparative Biochemistry and Physiology Part A: Physiology 76(1), 109–113.

Ruiz, G., Rosenmann, M., Veloso, A., 1989. Altitudinal distribution and blood values in the toad, *Bufo spinulosus* Wiegmann. Comparative biochemistry and physiology A, Comparative physiology 94(4), 643–646.

Saint Girons, M.C., 1970. Morphology of the circulating blood cells. Biology of the Reptilia 3(2), 73–92.

Shutler, D., Marcogliese, D.J., 2011. Leukocyte profiles of northern leopard frogs, *Lithobates pipiens*, exposed to pesticides and hematozoa in agricultural wetlands. Copeia 2011(2), 301–307.

Smith, H.M., 1925. Cell size and metabolic activity in Amphibia. The Biological Bulletin 48(5), 347–378.

Storer, T.I., Usinger, R.L., Stebbins, R.C., Nybakken, J.W., 1957. General zoology, McGraw-Hill.

Szarski, H., Czopek, G., 1966. Erythrocyte diameter in some amphibians and reptiles. Bulletin De L Academie Polonaise Des Sciences-Serie Des Sciences Biologiques 14(6), 433.

Tavares-Dias, M., De Moraes, F.R., Onaka, E.M., Rezende, P.C.B., 2007. Changes in blood parameters of hybrid tambacu fish parasitized by *Dolops carvalhoi* (Crustacea, Branchiura), a fish louse. Veterinarski arhiv 77(4), 355–363.

Thrall, M.A., Weiser, G., Allison, R., Campbell, T., 2012. Veterinary hematology and clinical chemistry. John Wiley & Sons.

Tierney, K., Farrell, A., Kennedy, C., 2004. The differential leucocyte landscape of four teleosts: juvenile *Oncorhynchus kisutch, Clupea pallasi, Culaea inconstans* and *Pimephales promelas*. Journal of Fish Biology 65(4), 906–919.

Vasaruchapong, T., Disarapong, P., Chulasugandha, P., Khow, O., Chanhome, L., Chiobamroongkiat, M., Chaiyabutr, N., Sitprija, V., 2014. Comparative studies on hematological and plasma biochemical parameters in different types of venomous snakes in Thailand. Comparative Clinical Pathology 23(4), 955–959.

Vernberg, F.J., 1955. Hematological studies on salamanders in relation to their ecology. Herpetologica 11(2), 129–133.

Weber, R.E., 2007. High-altitude adaptations in vertebrate hemoglobins. Respiratory physiology & neurobiology 158(2–3), 132–142.

Wojtaszek, J., Baranowska, M., Glubiak, M., Dzugaj, A., 1997. Circulating blood parameters of the water frog, *Rana esculenta* L. at pre-wintering stage. Zoologica Poloniae 42, 1–4.

Wright, K.N., Whitaker, B.R., 2001. Amphibian medicine and captive husbandry. Krieger Publication Company, 1 edition, 570p.